Optimizing Three-Dimensional Bioprinting for Cell Culture Scaffolds

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INTRODUCTION

Bioprinting Leukemia • Cellink BioX printer & bioinks • A type of blood cancer that • .stl from MAL originates within bone marrow • About 40% of Leukemia patients experience relapse after bone marrow transplant treatment, which has a high mortality rate¹ **Trabecular Bone** Spongy bone that houses the Figure 2. Trabecular Bone Gyroid Structure .stl File bone marrow • Print parameters changed: • Red blood cells, white blood cells, • speed, nozzle gauge, layer and platelets are formed² • Difficult to study in vivo due to the density location and type of tissue • G-Code Ink Dilution and Culture Cortical (hard) bone rabecular Conditions (media, PBS, Periosteum (spongy) bone (membrane covering bone) incubation) Articular cartilage **Testing and Characterization** • Ink testing: filament test, stack Marrow Blood

Figure 1. Trabecular Bone Diagram³

Medullary cavity

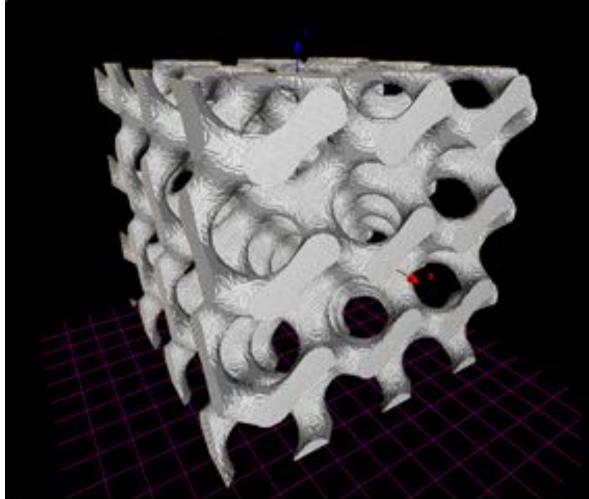
Epiphyseal plate

OBJECTIVE

vessels

Optimize and print a biomimetic trabecular bone scaffold to study cell interactions for improving leukemia treatment

METHODS



height, extrusion pressure, infill

test, layer height test⁴ • Z-stack pore size measurements 300-600 µm pore size for healthy trabecular bone⁵

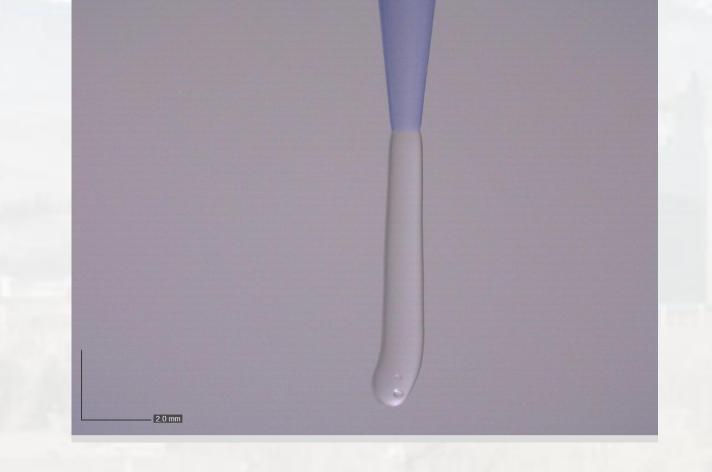


Figure 3. Bioink Filament Test (2mm scale bar)



RESULTS • Average short diameter of pores: 821.33 µm • Average long diameter of pores: 1299.01 µm • PBS destroys chemical cross linked prints*

Figure 5. Z-stack Image of

Figure 4. 3D Printed Bone Scaffold

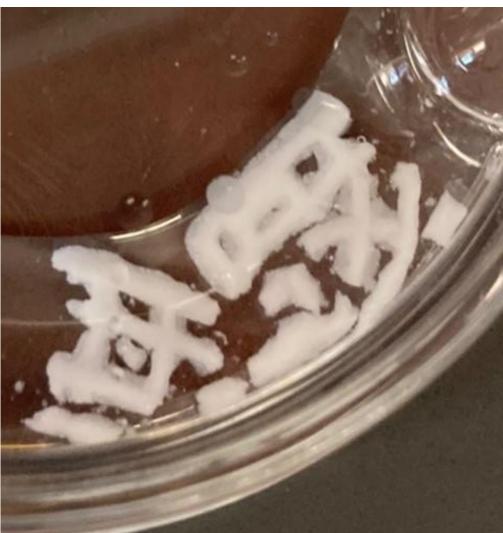


Figure 6. Dissolved Scaffold in PBS

 Best resolution with low pressure and low speed • Pore sizes are 135.6% bigger than actual* Diluted inks survived PBS soak, undiluted survived media soak

structure.

FUTURE WORK Incorporating additives • Mixing in cells Improving optimization protocols Ink testing Observing structural integrity for long-term culture conditions

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References

Barrett AJ, Battiwalla M. Relapse after allogeneic stem cell transplantation. Expert Rev Hematol. 2010 nstitute. Definition of bone marrow. Retrieved from A cross section of a human long bone SVG file by Pbroks13, distributed under a Creative Commons Attribution

- bility and Rheology







CONCLUSIONS

From the work done, the following conclusions were made:

1.The pore sizes measured may be closer to the appropriate size than it seems due to the structure of the scaffold and the measurement method. 2. The ion exchange that occurs between the crosslinking agent and the PBS may only occur at higher temperatures.

3. In order to increase resolution, smaller nozzle size may be necessary. 4. Incorporating supports and printing directly into the crosslinking agent has the possibility to provide a better

Acknowledgements

3.0 Unported license. Retrieved from https://commons.wikimedia.org/w/index.php?curid=5188772 O'Connell, C., et al. (2021). Characterizing Bioinks for Extrusion Bioprinting: Printability and Rheology. Method in molecular biology. 2140:111-133 https://www.researchgate.net/publication/340110752 Characterizing Bioinks for Extrusion Bioprinting Printa

Lee, S., Porter, M., Wasko, S. et al. Potential Bone Replacement Materials Prepared by Two Methods. MRS Online Proceedings Library 1418, 177–188 (2012). https://doi.org/10.1557/opl.2012.671

